

ANTIOXIDANT ACTIVITY OF MARINE RED ALGAE – *PORTIERIA HORNEMANNII* Louis Cojandaraj^{1#}, Surya Prabha U.² and Mary Elizabeth Shyamala³

^{1#} Department of Medical Laboratory sciences, Lovely professional University, Phagwara, Punjab, India. ² Department of Advanced Zoology and Biotechnology, Loyola College, Chennai, Tamilnadu, India

³ Department of Commerce, Holy Cross College, Trichy, Tamilnadu, India.

[#]Email address of Corresponding Author

louis.2333@lpu.co.in

Abstract

An *in vitro* evaluation study was conducted to assess the antioxidant capacity of the red algae *Portieria hornemannii* by using DPPH, Reducing power, ABTS radical scavenging activity and Superoxide scavenging activity.

Crude extracts (acetone, ethyl acetate, methanol and chloroform) was prepared by Soxhlet extraction and tested for the presence of active antioxidants using DPPH assay, Reducing power, ABTS assay and Superoxide scavenging activity and was statistically analysed with Tukey-HSD Homogenous subsets.

From the results the following inference can be obtained, in DPPH assay the ethyl acetate extract and methanol extract shows highest inhibition absorbance of 0.472 ± 0.002 and 0.484 ± 0.001 respectively. In reducing power assay, among the four extracts the chloroform extract shows absorbance of 0.198 ± 0.002 at $2.5 \ \mu g$, 0.297 ± 0.001 at $5 \ \mu g$, 0.305 ± 0.00 at $7.5 \ \mu g$ and 0.333 ± 0.002 at $10 \ \mu g$ respectively. In ABTS assay, the Ethyl acetate extract and chloroform extract shows highest inhibition absorbance of 0.435 ± 0.0010 and 0.488 ± 0.0025 . In Superoxide scavenging activity, the acetonic and methanolic extract shows 0.128 ± 0.001 and 0.138 ± 0.009 . The one-way ANOVA results of all the four assays showed a significance value of p < 0.01 which makes them highly significant.

The results prove the red algae *Portieria hornemannii* is a potential source as antioxidants against the reactive oxygen species which obstructs the cell metabolism and physiological activities.

Keywords: Portieria hornemannii, Anti-oxidant activity, Macro algae, Seaweed.

Introduction

Bioactive molecules from natural sources are attaining thrust in the field of natural product discovery. The terrestrial plants are being widely used while marine plants such as seaweeds are less exploited for the bioactive molecule's discovery. Hudson et al. (1998). In contrast to terrestrial vegetation, the macro algae produce bioactive metabolites in response to ecological pressures such as competition for space and ability to reproduce constitute valuable sources for drug development. De Vries & Beart (1995). In the past decade, the search for natural antioxidant compounds has gained considerable attention and the number of publications on antioxidants and oxidative stress has nearly quadrupled Huang, Ou & Prior (2005). Antioxidant compounds play an important role against various diseases like chronic inflammation, atherosclerosis, cancer and cardiovascular disorders. Kohen & Nyska (2002), which explains their considerable commercial potential in medicine, food production and the cosmetic industry. Moreover, interest in employing antioxidants from natural sources is considerably enhanced by consumer preference for natural products and concern about the potential toxic effects of synthetic antioxidants. Safer & Al-Nughamish (1999). Marine algae, like other photosynthesizing plants, are exposed to a combination of light and oxygen that leads to the formation of free radicals and other strong oxidizing agents. However, the absence of oxidative damage in the structural components of macro algae (i.e., polyunsaturated fatty acids) and their stability to oxidation during storage suggest that their cells have protective antioxidative defence systems. Fujimoto (1990); Matsukawa et al. (1997). In fact, algae have protective enzymes (superoxide dismutase, peroxidase, glutathione reductase, catalase) and antioxidative molecules (phlorotannins, ascorbic acid, tocopherols, carotenoids, phospholipids, chlorophyll related compounds, bromophenols, catechins, mycosporine-like amino acids, polysaccharides, etc.) which are similar to those of vascular plants. Fujimoto (1990); Le Tutour *et al.* (1998); Rupérez, Ahrazem & Leal (2002); Yuan & Song (2005).

This study reveals the antioxidant aspects exhibited by the marine red algae. Portieria hornemannii is a small red marine algal species which is widely distributed in tropical and subtropical water bodies of the Pacific and Indian Ocean. Guirv (2010)Portieria belongs to the family Rhizophyllidaceae. The family Rhizophyllidaceae includes 4 genera Contarinia, Ochtodes, Nesophila and Portieria. The geographical distribution of the species belonging to the genera is interesting and exclusive. Saunders, Chiovitti & Kraft (2004); Krayesky et al. (2009); Verbruggen et al. (2010). The present study was designed to investigate the antioxidant activity of Portieria hornemannii.

Materials and Methods

Collection of sample

Healthy and matured seaweeds were handpicked from the rocky intertidal region at the coastal region of Mandapam 9°28"31"0°N and 79°15"7"11°E, processed and stored at the laboratory. Y\ild\ir\im *et al.* (2000) The crude metabolites from the sample were extracted using Soxhlet extraction method using solvents like acetone, chloroform, ethyl acetate and methanol; the crude metabolites were exhibited for the antioxidant activity of *Portieria hornemannii*.

Anti-Oxidant Activity

DPPH Radical Scavenging Activity : DPPH (2, 2-diphenyl-2-picryl hydrazyl hydrate) is a free radical and it is relatively stable and a widely accepted a widely accepted radical for the estimation of radical scavenging activity. The reaction of reducing agents reducing agents with the free DPPH radical depends upon the electron taken up and the stoichiometric degradation of the color solution. Sala *et al.* (2002). DPPH Radical Scavenging Activity was carried determined according to the method of Molyneux and others, (2004).

Reducing Power : Reducing power of the extracts was evaluated according to the method of Oyaizu (1986) and Yen & Chen (1995) Potassium Ferro cyanide reacts with Fe^{3+} ions in the presence of an antioxidant agent and form a Prussian blue color which leads to the conversion of blue color to yellow color in the presence of antioxidant agent and hence the greater the reducing power of the compound greater the absorbance at 700nm. Zou *et al.* (2008)

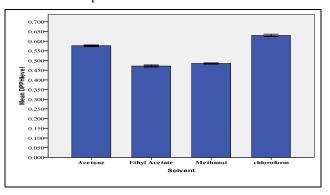
ABTS Radical Scavenging Activity : The antioxidant effect of the extracts was studied by ABTS (2, 2, –azino-bis-3-ethyl benzothiazoline -6-sulphonic acid) radical cationide colorization assay according to the method of Shirwaikar, Prabhu & Punitha (2006).

Superoxide Scavenging Activity : The superoxide scavenging ability of the extracts was assessed by the method of Winterbourn *et al.* (1975).

Statistical Analysis : The data collected were analyzed through statistical package for social studies version 17 (SPSS 17.0 ver.). The data were expressed as mean \pm SD statistical significance of variance was evaluated with one-way ANOVA, Two-way ANOVA with Tukey Multiple comparison. If the significance of variance i.e. P value (given under the head sign.is less than 0.05 (p < 0.05) the difference between the experimental conditions was considered significant. Regression and descriptive analysis were also carried out. The results obtained was also represented graphically.

Results and Discussion DPPH Radical Scavenging Activity

These radical comprises of highly reactive molecules or atoms which are unstable due to the presence of single electron or the presence of unbalanced electrons. Even though due to the presence of unbalanced electrons these radicals can be used for the normal cellular function under the physiological concentration, but the interruption in the function or the damage in the cellular metabolites such as lipids proteins and also in cellular components like nucleotide can occur due to the presence of excessive free radicals Zheng & Wang (2001). DPPH (2, 2-diphenyl-2picryl hydrazyl hydrate) Radical Scavenging Activity of the extracts (acetone, ethyl acetate, methanol and chloroform) was determined according to the method of Molyneux & others (2004). The mean \pm SD value were 0.576 \pm 0.001, $0.472 \pm 0.002, 0.484 \pm 0.001, 0.629 \pm 0.002$ (Graph 1). Higher level of DPPH was observed in the chloroformic extracts followed by the acetonic, methanolic and finally ethyl acetate extracts. From the results it was observed that chloroform extracts have high DPPH Radical Scavenging Activity compared with other extracts. The one-way analysis of variance showed that there is a significant (P<0.05)difference between the extract's treatments (Table 1). The results of Tukey-HSD Homogenous subsets formation showed that there are four subsets for the DPPH Radical Scavenging Activity (Table 2). The free radical scavenging activity of these antioxidants can protect from serious molecular and cellular damage and inhibit the process of several chronic disease and also lipid peroxidation in the food. Han et al. (2010) studied that the extracts of seaweeds pose anti-oxidant activity through DPPH radical scavenging activity and observed that the brown seaweed Cendaria pinnatifida expressed a good antioxidant activity through radical scavenging activity. Yan et al. (1999); Wang, Jonsdottir & Ólafsdóttir (2009) reported that higher amounts of polyphenols through DPPH radical scavenging activity and confined that brown algae exhibit more antioxidant activity than red and green algae. But in contrast to that Chandini, Ganesan & Bhaskar (2008) described in her studies that brown algae possess lower levels of DPPH radical scavenging activity ranging from17.79 to 23.16% at an extract concentration of 1000 µg/ml Duan et al. (2006) also discussed the crude extract of red alga, Polysiphonia urceolata exhibited a good DPPH radical scavenging activity. Hence DPPH a free radical can be extensively used as a free radical to evaluate the free radical scavenging activities of compounds.



Graph 1: Mean DPPH level of Portieria hornemannii

Table 1: One-way ANOVA - DPPH of Portieria hornemannii

		Sum of Squares	Df	Mean Square	F	Sig.
DPPH level	Between Groups	0.051	3	0.017	4869.778	0.000
	Within Groups	0.000	8	0.000		
	Total	0.051	11			

Table 2: Multiple Comparison	– DPPH of <i>Portieria hornemannii</i>
------------------------------	--

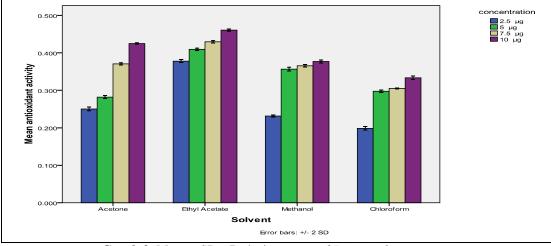
Tukey HSDª	Colvert	Solvent N	Subset for $alpha = 0.05$					
	Solvent	IN	1	2	3	4		
	Ethyl Acetate	3	.47200					
	Methanol	3		.48467				
115D	Acetone	3			.57633			
	chloroform	3				.62967		
	Sig.		1.000	1.000	1.000	1.000		
		Means for group	os in homogeneous	subsets are displaye	d.			
		a. Uses Ha	rmonic Mean Sam	ble Size = 3.000 .				

Reducing Power

The anti-oxidant potential activity of the compound can be indicated by the presence of reducing power of the compound. Lin *et al.* (2009) Reducing power of the extracts was evaluated according to the method of Oyaizu (1986); Yen & Chen (1995). The mean \pm SD value of the reducing power of the extracts (acetone, ethyl acetate, methanol and chloroform) at the concentration of 2.5 µg were 0.250, 0.378, 0.231, 0.198 and at the concentration of 5 µg were 0.282, 0.409, 0.356, 0.297 and at the concentration of 7.5 µg were 0.370, 0.429, 0.365, 0.305 and at the concentration of 10 µg were 0.425, 0.460, 0.377, 0.333. From the results it was observed that ethyl acetate extracts have high reducing power compared with other extracts (Graph 2). The two-way analysis of variance showed that there is a significant (P<0.05) difference between the extract's treatments (Table 3). The multiple comparison test Tukey-HSD (Homogenous subsets) also showed that there is a significant (P<0.05) difference between the extracts treatments (Table 4). Jiménez-Escrig *et al.* (2001) observed that red algae showed higher reducing power than brown algae due to the presence of polyphenols which is higher in red seaweeds comparing with green and brown seaweeds. Reducing power is exhibited in Seaweeds is due to the presence of flavonoids and phenols. Hence the seaweeds possess antioxidant, anti-allergic, anti-inflammatory, anti-microbial, anti-cancer activity. The antioxidant compounds during the reducing power assay donate electrons and also immediately reduce the lipid peroxidation process through reduced oxidation by which it can act has primary and secondary antioxidants. Yen & Chen (1995)

 Table 3 : Two-way ANOVA – Reducing power of Portieria hornemannii

	Tests of Between	-Subjects I	Effects					
	Dependent Variable: antioxidant activity							
Source	Type III Sum of Squares	df	Mean Square	F	Sig.			
Solvent	.115	3	.038	11334.887	.000			
concentration	.119	3	.040	11793.043	.000			
solvent * concentration	.022	9	.002	733.323	.000			
Error	.000	32	3.37					
Total	5.870	48						
Corrected Total	.257	47						



Graph 2: Mean ± SD	 Reducing power of 	f <i>Portieria</i> l	hornemannii
--------------------	---------------------------------------	----------------------	-------------

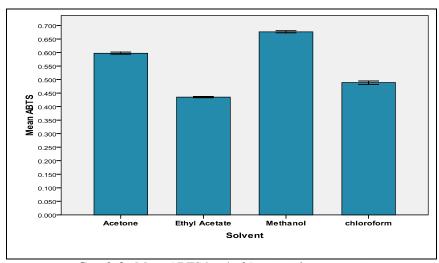
	Ν	Iultiple Comparisons of a	reducing power T	'ukey HSE)	
(I) solvent	(J) solvent	Mean Difference	C(L E	Sia	95% Confidence Interval	
(I) solvent	(J) solvent	(I-J)	Std. Error	Sig.	Lower Bound	Upper Bound
	Ethyl Acetate	08742*	.000750	.000	08945	08538
Acetone Vs	Methanol	00067	.000750	.811	00270	.00137
	Chloroform	$.04817^{*}$.000750	.000	.04613	.05020
	Acetone	$.08742^{*}$.000750	.000	.08538	.08945
Ethyl Acetate Vs	Methanol	.08675*	.000750	.000	.08472	.08878
	Chloroform	.13558*	.000750	.000	.13355	.13762
	Acetone	.00067	.000750	.811	00137	.00270
Methanol Vs	Ethyl Acetate	08675*	.000750	.000	08878	08472
	Chloroform	$.04883^{*}$.000750	.000	.04680	.05087
	Acetone	04817*	.000750	.000	05020	04613
Chloroform Vs	Ethyl Acetate	13558*	.000750	.000	13762	13355
	Methanol	04883*	.000750	.000	05087	04680
		Based on ob	served means.			
		The error term is Mean S	quare (Error) = 3.3	38E-006.		
		*. The mean difference is	significant at the .	05 level.		

Table 4: Multiple Comparison – Reducing power of Portieria hornemannii

ABTS Radical Scavenging Activity

ABTS radical scavenging activity is an indirect method for the determination of natural antioxidants these ABTS radical like the DPPH radical is also a free and rather stable ion in the absence of phenolic compound but it reacts energetically and gets converted into a non-colored form of ABTS with the help of H-atom donor like phenolic compound. Rauchová et al. (1995) The antioxidant effect for the extracts (acetone, ethyl acetate, methanol and chloroform) was studied by ABTS (2, 2, -azino-bis-3-ethyl benzothiazoline -6-sulphonic acid) radical cationide colorization assay according to the method of Shirwaikar et al. (2006). The mean \pm SD value were 0.597 \pm 0.0015, 0.435±0.001, 0.76±0.0015, 0.488±0.0025. From the results it was observed that methanol extracts have high ABTS Radical Scavenging Activity compared with other extracts (Graph 3). The one-way analysis of variance showed that there is a significant (P<0.05) difference between the extracts treatments (Table 5). The results of Tukey-HSD Homogenous subsets formation showed that there are four subsets for the ABTS Radical Scavenging Activity (Table 6). Sachindra et al. (2007) reported that the extracts obtained from the brown alga Turbinaria conoides and Padina tetrastomatica exhibited a higher ABTS radical scavenging activity compared to other four red seaweeds in its crude as well as fractionate extract derived from various solvents.. Bonnet, Camares & Veisseire (2000) observed 98%

inhibition of the ABTS molecule from the brown algae Padina minor. The results of the present study indicate that the methanolic extracts from Portieria hornemannii seaweed exhibited higher ABTS radical activity compared with other solvent extracts. However, the ABTS assay also possess certain limitations such as the competence of the sample to reacts with ABTS radical instead of inhibiting the oxidative process and decrease the reaction of many phenolics. Rauchová et al. (1995). Marinova & Yanishlieva (1997)., reported that the antioxidant activity is directly proportional to the types of solvent used for the extraction because the compounds obtained by the extraction depends upon the polarity difference and exhibit antioxidant potential at differing rates. Senthil Nathan, Kalaivani & Sehoon, (2006) reported that the ethanolic extract exhibited the maximum antioxidant activity which revealed that the maximum antioxidant activity was obtained from the polar solvents like ethanol than with extracts obtained from non-polar solvents.. Wang et al. (2009) reported that 70% aqueous acetone is more proficient to derive polyphenolic compounds from seaweeds rather than using water for extraction because the solubility of the Phenolic compounds is high in polar organic solvents than water. Gakunju et al. (1995) recommended that 70% aqueous acetone (v/v) was found to be the most efficient solvent and for the effective extraction of metabolites for the antioxidant activity the aqueous mixtures of methanol, ethanol and acetone are highly recommended.



Graph 3 : Mean ABTS level of *Portieria hornemannii* Table 5: One-way ANOVA – ABTS of *Portieria hornemannii*

		Sum of Squares	Df	Mean Square	\mathbf{F}	Sig.
ABTS	Between Groups	0.106	3	0.035	11756.769	0.000
	Within Groups	0.000	8	0.000		
	Total	0.106	11			

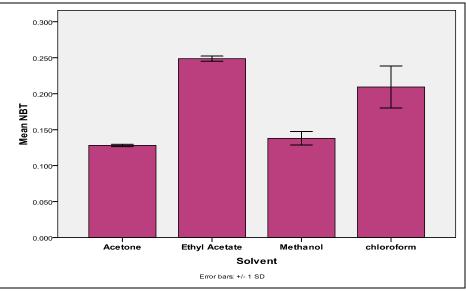
Table 6: Multiple Compariso	on – ABTS of <i>Portieria hornemannii</i>
-----------------------------	---

Tukey HSD ^a	Solvent	Solvent N	Subset for $alpha = 0.05$				
	Solvent	IN	1	2	3	4	
	Ethyl Acetate	3	.43500				
	chloroform	3		.48867			
	Acetone	3			.59733		
	Methanol	3				.67667	
	Sig.		1.000	1.000	1.000	1.000	
		Means for group	os in homogeneous s	ubsets are displayed.			
		a. Uses Ha	rmonic Mean Sampl	e Size = 3.000.			

Superoxide Scavenging Activity

The superoxide scavenging ability for the extracts (acetone, ethyl acetate, methanol and chloroform) was assessed by the method of Winterbourn et al. (1975). The mean ± SD value were 0.128±0.001, 0.248±0.003, 0.138±0.009, 0.209±0.0029. From the results it was observed that ethyl acetate extracts have high superoxide Scavenging Activity compared with other extracts (Graph 4). The oneway analysis of variance showed that there is a significant (P<0.05) difference between the extracts treatments (Table 7). The results of Tukey-HSD Homogenous subsets showed that there are two subsets formation for the superoxide Scavenging Activity. One subset for the acetone and methanol extracts and another subset for chloroform and ethyl acetate are tabulated (Table 8). Super oxide is one of the most important and a very effective free radical and it is

first radical to be generated in the cellular oxidation reaction and it acts as a precursor in the species utilizing oxygen to prevent the cellular damage by inhibiting the pathological activity of many diseases. Siriwardhana et al. (2003). obtained high values with Loboohora variegata and with some other species of Phaeophyta in the super oxide anion radical scavenging activity.. Athukorala et al. (2007) studied that the marine algae Grateloupia filficina obtained a higher super oxide anion radical scavenging activity and it was observed that the concentration range of 1mg/ml. Copernicia baileyana induced the production of superoxide anion radical instead of inhibiting it, in this context Zhang & Omaye (2001) stated that certain antioxidant like ascorbic acid and α tocopherol can act as pure oxidant depending upon the dose of the experimental conditions.



Graph 4: Mean Superoxide Scavenging Activity level of Portieria hornemannii

	Table 7. One-way ANOVA- Superovide Seavenging Activity level of Tortienta normemanni								
		Sum of Squares	Df	Mean Square	F	Sig.			
3T	Between Groups	0.030	3	0.010	41.372	0.000			

Table 7: One-v	vay ANOVA- Su	peroxide Scavenging	Activity level of	Portieria hornemannii

		Sum of Squares	Df	Mean Square	F	Sig.
NBT	Between Groups	0.030	3	0.010	41.372	0.000
	Within Groups	0.002	8	0.000		
	Total	0.032	11			

Table 8: Multiple Comparison - Superoxide Scavenging Activity level of Portieria hornemannii

Tukey HSD ^a	Solvent	N	Subset for alpha = 0.05			
			1			
	Acetone	3	.12800			
	Methanol	3	.13800			
	Chloroform	3		.20933		
	Ethyl Acetate	3		.24867		
	Sig.		.859	.058		
Means for g	groups in homogeneou	is subsets are disp	layed.	<u>.</u>		· · ·
a. Uses Har	monic Mean Sample	Size = 3.000.				

Conclusion

Herbal medicines are chief source of drug discovery as they are widely used for the control of pathogens and human related diseases. In the present investigation, it is proved the red algae Portieria hornemannii possess antioxidant activity and paves way in elucidating the key metabolite responsible for the activity against the reactive oxygen species. Further studies can be conducted on red algae Portieria hornemannii, marking them a significant source of metabolites for the cure of many fatal diseases like cancer, Alzheimer's disease.

Reference

Athukorala, Y.; Lee, K.-W.; Kim, S.-K. and Jeon, Y.-J. (2007). 'Anticoagulant activity of marine green and brown algae collected from Jeju Island in Korea', Bioresource Technology, 98(9): 1711–1716.

- Bonnet, M.; Camares, O. and Veisseire, P. (2000). 'Effects of zinc and influence of *Acremonium lolii* on growth parameters, chlorophyll a fluorescence and antioxidant enzyme activities of ryegrass (*Lolium perenne* L. cv Apollo)', Journal of Experimental Botany, 51(346): 945–953.
- Chandini, S.K.; Ganesan, P. and Bhaskar, N. (2008). '*In vitro* antioxidant activities of three selected brown seaweeds of India', Food chemistry, 107(2): 707–713.
- Duan, X.-J.; Zhang, W.-W.; Li, X.-M. and Wang, B.-G. (2006). 'Evaluation of antioxidant property of extract and fractions obtained from a red alga, Polysiphonia urceolata', Food chemistry, 95(1): 37–43.
- Fujimoto, K. (1990). 'Antioxidant activity of algal extracts', Introduction to applied phycology. SPB Academic Publishing, The Hague, 199–208.
- Gakunju, D.M.; Mberu, E.K.; Dossaji, S.F.; Gray, A.I.; Waigh, R.D.; Waterman, P.G. and Watkins, W.M. (1995). 'Potent antimalarial activity of the alkaloid nitidine, isolated from a Kenyan herbal remedy.', Antimicrobial agents and chemotherapy, 39(12): 2606– 2609.
- Guiry, M.D. (2010). 'AlgaeBase. World-wide electronic publication, National university of ireland, Galway', *http://www.algaebase.org/.*
- Han, J.W.; Jung, M.G.; Kim, M.J.; Yoon, K.S.; Lee, K.P. and Kim, G.H. (2010). 'Purification and characterization of a D-mannose specific lectin from the green marine alga, Bryopsis plumosa', Phycological research, 58(2): 143– 150.
- Huang, D.; Ou, B. and Prior, R.L. (2005). 'The chemistry behind antioxidant capacity assays', Journal of agricultural and food chemistry, 53(6): 1841–1856.
- Hudson, J.B.; Kim, J.H.; Lee, M.K.; DeWreede, R.E. and Hong, Y.K. (1998). 'Antiviral compounds in extracts of Korean seaweeds: Evidence for multiple activities', Journal of Applied Phycology, 10(5): 427.
- Jiménez-Escrig, A.; Jiménez-Jiménez, I.; Pulido, R. and Saura-Calixto, F. (2001). 'Antioxidant activity of fresh and processed edible seaweeds', Journal of the Science of Food and Agriculture, 81(5): 530–534.
- Kohen, R. and Nyska, A. (2002). 'Invited review: Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification', Toxicologic pathology, 30(6): 620–650.
- Krayesky, D.M.; Norris, J.N.; Gabrielson, P.W.; Gabriel, D. and Fredericq, S. (2009). 'A new order of red algae based on the Peyssonneliaceae, with an evaluation of the ordinal classification of the Florideophyceae (Rhodophyta)', Proceedings of the Biological Society of Washington, 122(3): 364–391.
- Lin, Ap.; Wang, C.; Qiao, H.; Pan, G.; Wang, G.; Song, L.; Wang, Z.; Sun, S. and Zhou, B. (2009). 'Study on the photosynthetic performances of *Enteromorpha prolifera* collected from the surface and bottom of the sea of Qingdao sea area', Chinese Science Bulletin, 54(3): 399–404.
- Marinova, E.M. and Yanishlieva, N.V. (1997). 'Antioxidative activity of extracts from selected species of the family Lamiaceae in sunflower oil', Food Chemistry, 58(3), 245–248.
- Matsukawa, R.; Dubinsky, Z.; Kishimoto, E.; Masaki, K.; Masuda, Y.; Takeuchi, T.; Chihara, M.; Yamamoto, Y.;

Niki, E. and Karube, I. (1997). 'A comparison of screening methods for antioxidant activity in seaweeds', Journal of Applied Phycology, 9(1): 29.

- Molyneux, P. and others, (2004). 'The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity', Songklanakarin J. sci. technol, 26(2): 211–219.
- Oyaizu, M. (1986). 'Studies on products of browning reaction', The Japanese journal of nutrition and dietetics, 44(6): 307–315.
- Rauchová, H.; Ledvinková, J.; Kalous, M. and Drahota, Z. (1995). 'The effect of lipid peroxidation on the activity of various membrane-bound ATPases in rat kidney', The international journal of biochemistry & cell biology, 27(3): 251–255.
- Rupérez, P.; Ahrazem, O. and Leal, J.A. (2002). 'Potential antioxidant capacity of sulfated polysaccharides from the edible marine brown seaweed *Fucus vesiculosus*', Journal of agricultural and food chemistry, 50(4): 840–845.
- Sachindra, N.M.; Sato, E.; Maeda, H.; Hosokawa, M.; Niwano, Y.; Kohno, M. and Miyashita, K. (2007). 'Radical scavenging and singlet oxygen quenching activity of marine carotenoid fucoxanthin and its metabolites', Journal of agricultural and food chemistry, 55(21): 8516–8522.
- Safer, A.M. and Al-Nughamish, A.J. (1999). 'Hepatotoxicity induced by the anti-oxidant food additive, butylated hydroxytoluene (BHT), in rats: an electron microscopical study', Histology and histopathology, 14(2): 391–406.
- Sala, A.; Recio, M. del C.; Giner, R.M.; Máñez, S.; Tournier, H.; Schinella, G. and Rios, J.-L. (2002). 'Antiinflammatory and antioxidant properties of Helichrysum italicum', Journal of Pharmacy and Pharmacology, 54(3): 365–371.
- Saunders, G.W.; Chiovitti, A. and Kraft, G.T. (2004). 'Smallsubunit rDNA sequences from representatives of selected families of the Gigartinales and Rhodymeniales (Rhodophyta). 3. Delineating the Gigartinales sensu stricto', Canadian journal of botany, 82(1): 43–74.
- Senthil, N.S.; Kalaivani, K. and Sehoon, K. (2006). 'Effects of *Dysoxylum malabaricum* Bedd.(Meliaceae) extract on the malarial vector Anopheles stephensi Liston (Diptera: Culicidae)', Bioresource Technology, 97(16): 2077–2083.
- Shirwaikar, A.; Prabhu, K.S. and Punitha, I.S.R. (2006). 'In vitro antioxidant studies of Sphaeranthus indicus (Linn)'.
- Siriwardhana, N.; Lee, K.-W.; Jeon, Y.-J.; Kim, S.-H. and Haw, J.-W. (2003). 'Antioxidant activity of Hizikia fusiformis on reactive oxygen species scavenging and lipid peroxidation inhibition', Food Science and Technology International, 9(5): 339–346.
- Tutour, B. Le; Benslimane, F.; Gouleau, M.P.; Gouygou, J.P.; Saadan, B. and Quemeneur, F. (1998).
 'Antioxidant and pro-oxidant activities of the brown algae, *Laminaria digitata, Himanthalia elongata, Fucus vesiculosus, Fucus serratus* and *Ascophyllum nodosum*', Journal of Applied Phycology, 10(2): 121–129.
- Verbruggen, H.; Maggs, C.A.; Saunders, G.W.; Gall, L. Le; Yoon, H.S. and Clerck, O. De (2010). 'Data mining approach identifies research priorities and data

requirements for resolving the red algal tree of life', BMC evolutionary biology, 10(1): 16.

- Vries, D.J. De and Beart, P.M. (1995). 'Fishing for drugs from the sea: status and strategies', Trends in pharmacological sciences, 16(8): 275–279.
- Wang, T.; Jonsdottir, R. and Ólafsdóttir, G. (2009). 'Total phenolic compounds, radical scavenging and metal chelation of extracts from Icelandic seaweeds', Food chemistry, 116(1): 240–248.
- Winterbourn, C.C.; Hawkins, R.E.; Brian, M. and Carrell, R.W. (1975). 'The estimation of red cell superoxide dismutase activity', The Journal of laboratory and clinical medicine, 85(2): 337–341.
- Y\ild\ir\im, A.; Mavi, A.; Oktay, M.; Kara, A.A.; Algur, Ö.F. and Bilalolu, V. (2000). 'Comparison of antioxidant and antimicrobial activities of Tilia (*Tilia argentea* Desf ex DC), sage (*Salvia triloba* L.), and Black tea (*Camellia sinensis*) extracts', Journal of Agricultural and food chemistry, 48(10): 5030–5034.
- Yan, X.; Chuda, Y.; Suzuki, M. and Nagata, T. (1999). 'Fucoxanthin as the major antioxidant in *Hijikia fusiformis*, a common edible seaweed', Bioscience,

biotechnology, and biochemistry, 63(3): 605–607.

- Yen, G.-C. and Chen, H.-Y. (1995). 'Antioxidant activity of various tea extracts in relation to their antimutagenicity', Journal of agricultural and food chemistry, 43(1): 27–32.
- Yuan, H. and Song, J. (2005). 'Preparation, structural characterization and *in vitro* antitumor activity of kappa-carrageenan oligosaccharide fraction from Kappaphycus striatum', Journal of Applied Phycology, 17(1): 7–13.
- Zhang, P. and Omaye, S.T. (2001). 'Antioxidant and prooxidant roles for $\beta\beta$ -carotene, α -tocopherol and ascorbic acid in human lung cells', *Toxicology in vitro*, 15(1): 13–24.
- Zheng, W. and Wang, S.Y. (2001). 'Antioxidant activity and phenolic compounds in selected herbs', Journal of Agricultural and Food chemistry, 49(11): 5165–5170.
- Zou, Y.; Qian, Z.-J.; Li, Y.; Kim, M.-M.; Lee, S.-H. and Kim, S.-K. (2008). 'Antioxidant effects of phlorotannins isolated from *Ishige okamurae* in free radical mediated oxidative systems', Journal of Agricultural and Food Chemistry, 56(16): 7001–7009.